



The Role of ADMA in Various Diseases

Sofna Banjarnahor,¹ Reny Damayanti²

¹Pusat Penelitian Kimia – Badan Riset dan Inovasi Nasional (BRIN), Kawasan PUSPIPTEK Serpong, Tangerang Selatan, Indonesia,

²Laboratorium Patologi Klinik, Rumah Sakit Pusat Angkatan Darat (RSPAD) Gatot Soebroto, Jakarta, Indonesia

ABSTRACT

Asymmetric dimethylarginine (ADMA) is a competitive inhibitor of nitric oxide synthase (NOS). Numerous studies have discovered a correlation between high plasma ADMA levels and the development of multiple diseases, including cardiovascular disease, chronic renal disease, diabetes mellitus, liver diseases, preeclampsia, and COVID-19. In addition, ADMA has been established as an independent cardiovascular risk factor. However, the functional interplay between ADMA and nitric oxide (NO)-mediated pathways is poorly understood, leaving the distinction between “risk factor” and “risk marker” unclear. In this review, we provide insights into the current state of knowledge regarding the pathophysiological role of ADMA in various diseases, the relationship between ADMA and endothelial dysfunction, the current analytical techniques used to determine ADMA in body fluids, and the benefits of ADMA-lowering drugs for the prevention of diseases in humans.

Keywords: ADMA, cardiovascular disorders, nitric oxide, risk marker

ABSTRAK

Asymmetric dimethylarginine (ADMA) adalah penghambat kompetitif *nitric oxide synthase* (NOS). Banyak penelitian menemukan hubungan antara peningkatan kadar plasma ADMA dan perkembangan berbagai penyakit, termasuk penyakit kardiovaskular, penyakit ginjal kronik, diabetes melitus, penyakit hati, preeklampsia, dan COVID-19. ADMA juga diidentifikasi sebagai faktor risiko independen untuk penyakit kardiovaskular. Namun, interaksi fungsional antara ADMA dan *nitric oxide* (NO) belum begitu dipahami, yang menyebabkan tidak jelasnya batasan antara “faktor risiko” dan “penanda risiko”. Tulisan ini membahas pengetahuan terbaru tentang peran patofisiologi ADMA dalam berbagai penyakit, hubungan antara ADMA dan disfungsi endotelial, teknik analisis terkini untuk pemeriksaan kadar ADMA dalam cairan tubuh, serta manfaat obat penurun ADMA untuk pencegahan penyakit pada manusia. **Sofna Banjarnahor, Reny Damayanti. Peran ADMA pada Berbagai Penyakit.**

Kata kunci: ADMA, penyakit kardiovaskular, *nitric oxide*, penanda risiko

INTRODUCTION

The accumulation of asymmetric dimethylarginine (ADMA) has been related to several vascular diseases. Vallance, *et al*, were the first to suggest that ADMA may have a pathophysiological role as a competitive inhibitor of nitric oxide synthase (NOS). In addition, they found an increase in methylarginine in the plasma of patients with end-stage renal failure.¹ Since then, ADMA has been discovered as an independent risk factor for cardiovascular events.² ADMA meets several criteria for a uremic toxin: it accumulates when the kidney fails, is a byproduct of protein metabolism, is a guanidino compound, and has the potential to affect numerous biological pathways disrupted in patients with chronic renal failure, such as the cardiovascular system, bone, and immune system.¹

Given the clinical importance of ADMA on vascular (i.e., endothelial) health, three studying ADMA metabolism, particularly its transport and clearance pathways, may provide a complete picture of how this metabolite exerts its harmful effects. Consequently, this study aims to provide clinicians with knowledge on ADMA and, where available, research linking ADMA to human diseases. The following section briefly outlines how ADMA is examined and if it may be utilized in clinical practice. Most importantly, lowering ADMA in individuals with ADMA-related diseases has benefits.

ADMA Metabolism and Transport

In 1970, Kakimoto and Akazawa isolated and characterized ADMA in human urine.⁴ They stated that ADMA was formed endogenously and was unaffected by diet. Using high-performance liquid chromatography (HPLC),⁵

ADMA may be quantified in human plasma.⁶ Protein arginine N-methyl-transferases (PRMT) generate ADMA by methylating protein-bound L-arginine post-translationally.⁷

ADMA is produced when PRMT methylates the guanidine group of L-arginine. Both types of PRMT catalyze the monomethylation of L-arginine to NG-monomethyl arginine (NMMA).⁷ Attachment of the second methyl group by PRMT types 1 or 2 generates different product-asymmetric (ADMA) and symmetric dimethylarginine (SDMA) - respectively.

Most ADMA is either degraded in the cytoplasm of the cell that made it or eliminated through the kidneys. Due to its physical features, ADMA, like other cationic amino acids, requires specific transport systems known as amino acid transporters. Two methods are known to transport cationic amino acids: y⁺ and y⁺L.

Alamat Korespondensi email: banjarnahorsofna@gmail.com



The $\gamma+$ system, known as cationic amino acid transporters (CATs), can transport ADMA and its analog molecule, L-arginine, across the cell membrane⁸ and to other cells.⁹ Whereas the γ +L system transports neutral and cationic amino acids.¹⁰

ADMA clearance by the kidney is mediated through the action of dimethylarginine dimethylaminohydrolases (DDAHs).¹¹ This enzymatic family in higher organisms consists of two isoforms encoded by genes on chromosomes 1 (DDAH-1) and 6 (DDAH-2), with distinct tissue-specific distributions but seemingly similar activities.¹² In addition, subsequent research has implicated the liver in the metabolism of dimethylarginines. Researchers have demonstrated that hepatocyte membranes contained abundant $\gamma+$ channels¹³ and high concentrations of DDAH.¹⁴ A recent study on patients undergoing hepatectomy confirmed the presence of ADMA in the liver.¹⁵ These metabolic studies have shown that the liver is essential for regulating plasma dimethylarginine concentrations.

CLINICAL OVERVIEW

The plasma concentration of ADMA in healthy adults ranges between 0.4 M and 1 M;¹⁶ it can increase to between 1.45 M and 4.0 M in some diseases. Given the growing understanding of the roles of ADMA in the pathogenesis of conditions, the following part focuses on disorders associated with elevated plasma levels of ADMA.

ADMA in Cardiovascular Diseases

Numerous investigations have established a strong correlation between increased ADMA levels and cardiovascular diseases. Kielstein, *et al*, observed that patients with hypertension had greater plasma levels of ADMA than normal-tension individuals.¹⁷ Elevated plasma ADMA levels have also been found in hypertensive youngsters.¹⁸ In addition, Wang, *et al*,¹⁹ and Surdacki, *et al*,²⁰ reported elevated plasma levels of ADMA in newly diagnosed and treated hypertension patients. In addition, high plasma ADMA in individuals with myocardial infarction or chronic left heart failure is one of the strongest predictors of mortality.⁹

ADMA levels are elevated in patients with heart failure,²¹⁻²³ and it has been demonstrated

that ADMA reduces ventricular contraction and heart rate.²⁴ According to Kielstein, *et al*,²⁵ exogenous ADMA increases systemic vascular resistance and mean arterial pressure while lowering cardiac output in the male group. In addition, they observed that ADMA infusion decreased renal circulation and salt reabsorption related to dosage. ADMA may contribute to the pathogenesis of cardiovascular diseases via two distinct mechanisms. On the one hand, ADMA inhibits endothelial nitric oxide synthase (eNOS) activity, resulting in vasoconstriction of blood vessels.^{24,26} On the other hand, ADMA suppresses nitric oxide (NO) synthesis in the kidneys, resulting in reduced salt excretion.²⁷⁻²⁹

ADMA in Chronic Kidney Diseases

High ADMA levels have also been found in human patients with kidney diseases and have been identified as a predictive factor in chronic kidney disease (CKD) patients.³⁰ Vallance, *et al*, were the first to show increased plasma levels of ADMA in CKD patients.¹ A recent meta-analysis of 18 studies with a total of 2136 patients with varying stages of renal failure revealed that plasma concentrations of ADMA were up to 3.4 times higher in renal insufficiency patients on dialysis compared to healthy control groups.³¹ However, it is doubtful whether the elevated ADMA levels result from a pathogenic process or an inflammatory byproduct.

According to reports, the kidney is the primary organ responsible for ADMA elimination.³² The greatest expression and activity of the ADMA breakdown enzyme DDAH¹ is found in the kidneys.³³ ADMA accumulation can occur from reduced DDAH1 activity^{34,35} via enhanced DDAH¹ degradation by post-translational modifications such as DDAH¹ protein oxidation^{36,37} or DDAH¹ gene loss-of-function mutations.³⁸ Therefore, if the kidney fails, ADMA will accumulate, making it a predictor for renal failure. Increased ADMA, on the other hand, may affect various renal functions, such as tubuloglomerular feedback (TGF) response, tubular sodium and proton transport, and renal sympathetic nerve activity,³⁹ making ADMA a causative factor of kidney diseases.

ADMA in Diabetes Mellitus

High levels of plasma ADMA are also found in diabetic patients who develop microvascular complications in the retina,⁴⁰ nerves,⁴¹ and

kidneys.⁴² A meta-analysis found that ADMA might have a role in the genesis of diabetic microvascular complications.⁴³ Lin, *et al*, observed a correlation between diabetic rat DDAH activity and ADMA levels.³⁷ When human endothelial cells (HMEC-1) and vascular smooth muscle cells were exposed to a high-glucose medium, DDAH activity was significantly reduced. Hyperglycemia-induced oxidative stress leads to ADMA accumulation, which may contribute to endothelial vasodilator dysfunction resulting from insufficient NO production. In addition, Lee, *et al*, demonstrated in a cross-sectional study that increased ADMA concentrations may also inhibit insulin-stimulated glucose absorption in skeletal muscle by altering insulin signaling.⁴⁴ Furthermore, they stated that ADMA was pathophysiologically significant as a potential biomarker of insulin resistance in skeletal muscle. Whether ADMA accumulation affects insulin signaling and insulin resistance in humans must be determined. These results reveal the possibility of a novel therapeutic target with clinical relevance for regulating energy and metabolic balance.

ADMA in Liver Diseases

Plasma ADMA concentrations have increased in individuals with hepatic impairment.⁴⁵ Notably, plasma ADMA levels are strongly correlated with the severity of hepatic dysfunction in patients with liver diseases of diverse etiologies.^{46,47} Individuals with alcoholic hepatitis,⁴⁸ liver cirrhosis,⁴⁹ and acute liver failure have elevated plasma ADMA levels.⁴⁵ Plasma ADMA levels are associated with hepatic impairment in cirrhosis and are much higher in individuals with decompensated cirrhosis than in those with compensated cirrhosis.⁴⁸ Due to the increased expression of the ADMA-degrading enzyme DDAH in the liver, the liver plays an indispensable role in ADMA elimination. Animal studies have indicated a negative correlation between serum ADMA concentrations and liver DDAH activity.⁵⁰ The significance of ADMA in liver disease is tied to its ability to induce hepatic malperfusion. Loss of liver function interferes with DDAH activity and the subsequent clearance of ADMA, resulting in a considerable rise in plasma ADMA levels and inhibition of NO generation, reducing hepatic flow and affecting other organs.⁴⁷



ADMA in Preeclampsia

Elevated ADMA levels in women with preeclamptic pregnancy have been demonstrated in many studies.^{51, 52} Plasma ADMA levels are increased when measured at the 23-25th week of pregnancy, even before the onset of preeclampsia,⁵³ similar to what Speer and colleagues found in their studies.⁵⁴ Pettersson, *et al*,⁵⁵ showed that plasma ADMA was higher in third-trimester preeclamptic patients (0.55 ± 0.02 mmol/l; $p < 0.05$) compared to normotensive pregnant controls (0.36 ± 0.01 mmol/l) of the same gestational length. A substantial difference was also observed between preeclamptic and normal pregnant women. In contrast, extensive studies involving Colombian women showed no significant difference in ADMA levels between normal pregnancy and preeclamptic pregnancy.⁵⁶ It has been suggested that the relative cardiovascular and infectious risk of a community may explain this leveling of ADMA-associated risk.⁵²

Whether ADMA has a role in normal pregnancies remains undetermined. Maeda, *et al*,⁵⁷ discovered that maternal concentrations of ADMA were lower in normal pregnancies than in non-pregnant women, indicating that ADMA is essential for vasodilation during pregnancy. Recently, Saarelainen, *et al*,⁵⁸ have cast doubt on the role of ADMA as a critical regulator of blood pressure in normal pregnancy. Their study demonstrated only subtle alterations of ADMA and no correlation between endothelium-dependent vasodilatation markers and maternal serum concentrations of ADMA or L-arginine. Similarly, Braekke, *et al*, found that maternal concentrations of ADMA and L-arginine were significantly higher in preeclamptic women than in healthy controls. They suggested that the effect of ADMA on NOS could be attenuated by high L-arginine, which obscures the previous suggestion of ADMA as a predominant factor of endothelial function in preeclampsia.⁵⁹

Interestingly, ADMA levels are elevated even before preeclampsia onset^{53,54} suggesting that ADMA could play a role in the development of preeclampsia and offers a potential role for ADMA as a novel risk marker for the early detection of high-risk pregnancy. These results suggest that ADMA influences preeclampsia via a complex mechanism. ADMA may be

elevated due to an excess of methylated arginine residues on proteins, the metabolism of ADMA by DDAH may be impaired, or elevated ADMA may come from underlying changes in renal function.

ADMA in COVID-19

Hanemann, *et al*, initially found a correlation between elevated plasma levels of ADMA and COVID-19 severity. They observed that ADMA might be used to identify hospitalized COVID-19 patients with a high probability of hospital mortality.⁶⁰ It is unknown how ADMA contributes to the pathophysiology of COVID-19, but decreased nitric oxide (NO) production by endothelial cells may play a role in COVID-19-associated morbidities. Impaired NO production may interfere with two key pathophysiological processes: vascular dysfunction and immune function.⁶¹

Endothelial cells infected with the SARS-CoV-2 virus develop endotheliopathy,⁶² perhaps resulting in endothelial dysfunction and NO deficiency due to cell tropism. According to Ueda, *et al*,⁶³ the ADMA-DDAH system regulates inflammation-mediated NO production via inducible NOS (iNOS). Expression of the third isoform, iNOS, is dependent on inflammatory mediators (i.e., cytokines, bacterial lipopolysaccharide, and interferons)⁶⁴ and virus infection,⁶⁵ and other pathophysiological stimuli.^{66,67}

Upon induction, iNOS can produce 1,000 times more NO than eNOS and persist until the enzyme is destroyed, which can take several hours due to the tight interaction between calmodulin and the enzyme.⁶⁸ iNOS-derived NO can lead to harmful effects not due to the direct actions of NO, but rather due to the abundance of NO available to react with superoxide radical (O_2^-), leading to the formation of the highly reactive (and harmful) radical, peroxynitrite (ONOO⁻), and further downstream, derivatives such as nitryl and hydroxyl peroxide.⁶⁹ These harmful effects have been linked to several human illnesses,⁷⁰ including COVID-19.⁶² A study demonstrated that plasma nitric oxide (NO) species were lower in COVID-19 patients than in healthy controls due to a decreased production or increased consumption of reactive oxygen species scavenging.⁷¹ Others have recommended monitoring nitrite/nitrate levels (NOx) and even supplementing COVID-19 with exogenous nitrate or nitrite.⁷²

Therefore, inflammation-induced oxidative stress, as seen in COVID-19, modifies the activity of the redox-sensitive ADMA-related enzymes PRMT and DDAH.⁷³ As a result, plasma ADMA levels increase, boosts the inhibition of eNOS and iNOS, and worsens the clinical condition.

ADMA-mediated Endothelial Dysfunction

The endothelium is essential for maintaining vascular tone and structure. Endothelial dysfunction is associated with cardiovascular, metabolic, and systemic or local inflammation risk factors.⁷⁴ Elevated blood levels of ADMA, an analog of the amino acid L-arginine, is one proposed mechanism for developing endothelial dysfunction.⁷⁵

As noted above, ADMA inhibits all three NOS isoforms for the formation of nitric oxide,¹ the most potent endogenous vasodilator and a critical signaling molecule for numerous molecular targets.⁷⁶ A high concentration of ADMA could therefore impair vascular function. ADMA causes an increase in renal vascular resistance and blood pressure in rats.⁷⁷ Additionally, local intraarterial infusion of ADMA has been reported to reduce forearm blood flow significantly.⁷⁸ Moreover, intravenous infusion of ADMA increased mean blood pressure by 6 percent and systemic vascular resistance by 24 percent while reducing the effect of exercise on cardiac output (by 15 percent), heart rate, and vascular responsiveness.²⁴ These findings confirm its biological action *in vivo*.

Some have argued, however, that the concentration of ADMA in plasma, even in disease states, is too low to inhibit NOS effectively and that the average concentrations of L-arginine in cells should counteract the inhibitory effects of ADMA on NOS.⁷⁹ It should be highlighted that the interaction of ADMA with eNOS should be more comprehensive than just ADMA levels alone. Other cellular processes, such as those involving the transport and metabolism of ADMA and L-arginine, may be involved since L-arginine shares the same transporter with ADMA and is also the sole substrate for NO generation.⁸⁰

ADMA Measurement

It has been demonstrated that ADMA concentrations in the general population fall within a restricted range.⁸¹ Even when slightly increased, ADMA is associated with considerable cardiovascular risk.⁸² Consequently, accurate



testing is necessary. Several variables regulate ADMA plasma concentrations; age,⁸³ race,⁸⁴ and body mass are among these determinants.⁸⁴ The measurement method is an additional crucial element influencing the concentration of ADMA.⁸³ Numerous analytical techniques for measuring ADMA in plasma and urine have been described. Among them are the enzyme-linked immunosorbent assay (ELISA), high-performance liquid chromatography (HP-LC) with fluorescence detection, capillary electrophoresis, liquid chromatography coupled to mass spectrometry (LC-MS), and gas chromatography coupled to mass spectrometry (GC-MS).⁸⁵

Each technique has its advantages and disadvantages. The most critical aspect of ELISA is that it is helpful in both routine diagnostic practice and clinical investigations. It is also very efficient and sensitive because it only measures one analyte simultaneously. However, compared to HPLC and MS-based methods, it has lower sensitivity for detecting ADMA in circulation.⁸⁶ The reduction in analysis time from a few hours to 30 to 60 minutes is a significant advantage of HPLC over other techniques. This method can also detect ADMA in biological fluid samples such as cell culture samples, tissue extracts, urine, plasma, and serum.⁸⁶ HPLC is now the most widely used method.⁸⁵ The HPLC-based method, on the other hand, cannot distinguish between ADMA and its structural analogs, such as symmetric dimethylarginine (SDMA) and L-arginine.⁸⁷ Mass spectrometry (MS)-based procedures provide a more precise determination of ADMA.⁸⁸ However, these methods require prohibitively expensive equipment for some facilities. Despite being established as a cardiovascular risk marker, plasma ADMA measurement is not yet standard in clinical practice.

ADMA-lowering Drugs

Is ADMA reduction beneficial? Extensive investigations have established causal links between ADMA and vascular function,⁷⁵ highlighting the potential advantages of a particular ADMA-reducing drug. No medications are currently available for ADMA-lowering treatment. The experimental data reveal that PRMT1, DDAH2, and CAT1 transporter modulate ADMA's intracellular and circulation amounts.⁸⁹

At least three potential targets for treatments in ADMA-mediated vascular disease are theorized to exist. The first option is to inhibit PRMT1 specifically. However, creating a selective inhibitor for PRMT1, an enzyme with significant sequence conservation during evolution, is difficult.⁹⁰ In addition, as PRMT is implicated in complicated cellular physiology, PRMT1 inhibition might be a scam with adverse side effects.⁹¹ Nonetheless, and several potent PRMT inhibitors require more experimental confirmation.⁹²

Second, recent advancements in understanding the control of DDAH activity may yield significant new therapeutic opportunities.³³ Compounds, such as aminoguanidine,⁹³ pioglitazone,⁹⁴ pravastatin,⁹⁵ probucol,⁹⁶ farnesoid X receptor agonists,⁹⁷ melatonin,⁹⁸ and vitamin E⁹⁹ have been found as potentially therapeutically significant stimulants of DDAH enzyme activity and expression, hence lowering ADMA levels. Recently, a small molecule developed by modifying DDAH with polyethyleneglycol (PEG)ylation reduced blood pressure and improved renal function through lowering ADMA levels in animal models.¹⁰⁰ However, more examination is required before these "bench" results may be effectively translated to the "bedside."

ADMA buildup can be altered by inhibiting

ADMA transport mediated by CAT1.¹⁰¹ Recent *in vitro* investigations of the effects of some commonly prescribed medicines have indicated that some of these substances interfere with ADMA transport,¹⁰² possibly changing extracellular and intracellular ADMA concentrations. However, it remains uncertain whether ADMA reduction treatment research conducted *in vitro* can be applied to humans. In addition, clinical research has investigated the impact of statins, angiotensin-converting enzyme inhibitors, angiotensin receptor blockers, metformin, hormone replacement therapy, fenofibrate, folate, and α -lipoic acid on the lowering of plasma ADMA levels.¹⁰³⁻¹⁰⁶ However, it is still debatable whether a 20% drop in plasma ADMA levels generated by these medicines is beneficial in treating various disorders.

CONCLUSION

After over 30 years of research, ADMA is becoming a therapeutically functional metabolite. Despite this, several crucial ADMA concerns still need to be addressed. Because ADMA is processed by endothelial cells, a rise in ADMA may be the outcome of endothelial dysfunction rather than its cause. Although the exact reason for higher ADMA levels in diseases is unknown, the outlined scenarios can explain the rise. Increased PRMT-mediated ADMA synthesis in response to stress; increased breakdown or protein turnover, including methylated arginine; decreased ADMA degradation due to decreased DDAH activity, or decreased ADMA excretion as a result of renal dysfunction. Improved ADMA detection methods that permit routine clinical sample analysis would greatly assist future research. It is strongly desired that further study be conducted on its relevance as a causal factor and treatment development target. Hence, more research has to be focused on ADMA to reach clinical applications.

REFERENCES

- Vallance P, Leone A, Calver A, Collier J, Moncada S. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* 1992;339(8793):572-5.
- Vallance P. Importance of asymmetrical dimethylarginine in cardiovascular risk. *The Lancet* 2001;358(9299):2096-7.
- Leiper J, Nandi M, Torondel B, Murray-Rust J, Malaki M, O'Hara B, et al. Disruption of methylarginine metabolism impairs vascular homeostasis. *Nat Med.* 2007;13(2):198-203.
- Kakimoto Y, Akazawa S. Isolation and identification of N-G,N-G- and N-G,N'-G-dimethyl-arginine, N-epsilon-mono-, di-, and trimethyllysine, and glucosylgalactosyl- and galactosyl-delta-hydroxylysine from human urine. *J Biol Chem.* 1970;245(21):5751-8.
- Nakajima T, Matsuoka Y, Kakimoto Y. Isolation and identification of N-G-monomethyl, N-G, N-G-dimethyl- and N-G,N' G-dimethylarginine from the hydrolysate of proteins of bovine brain. *Biochim Biophys Acta.* 1971;230(2):212-22.
- Park KS, Lee HW, Hong SY, Shin S, Kim S, Paik WK. Determination of methylated amino acids in human serum by high-performance liquid chromatography. *J Chromatogr.* 1988;440:225-30.
- Gary JD, Clarke S. RNA and protein interactions modulated by protein arginine methylation. *Prog Nucleic Acid Res Mol Biol.* 1998;61:65-131.



8. Closs EI, Basha FZ, Habermeier A, Förstermann U. Interference of L-arginine analogues with L-arginine transport mediated by the y^+ carrier hCAT-2B. *Nitric Oxide* 1997;1(1):65-73.
9. Liu X, Hou L, Xu D, Chen A, Yang L, Zhuang Y, et al. Effect of asymmetric dimethylarginine (ADMA) on heart failure development. *Nitric Oxide* 2016;54:73-81.
10. Mann GE, Yudilevich DL, Sobrevia L. Regulation of amino acid and glucose transporters in endothelial and smooth muscle cells. *Physiol Rev.* 2003;83(1):183-252.
11. McDermott JR. Studies on the catabolism of Ng-methylarginine, Ng, Ng-dimethylarginine and Ng, Ng-dimethylarginine in the rabbit. *Biochem J.* 1976;154(1):179-84.
12. Leiper JM, Santa Maria J, Chubb A, MacAllister RJ, Charles IG, Whitley GS, et al. Identification of two human dimethylarginine dimethylaminohydrolases with distinct tissue distributions and homology with microbial arginine deiminases. *Biochem J.* 1999;343 Pt 1(Pt 1):209-14.
13. Hattori Y, Kasai K, Gross SS. Cationic amino acid transporter gene expression in cultured vascular smooth muscle cells and in rats. *Am J Physiol.* 1999;276(6):2020-8.
14. Kimoto M, Whitley GS, Tsuji H, Ogawa T. Detection of NG,NG-dimethylarginine dimethylaminohydrolase in human tissues using a monoclonal antibody. *J Biochem.* 1995;117(2):237-8.
15. Siroen MP, van der Sijp JR, Teerlink T, van Schaik C, Nijveldt RJ, van Leeuwen PA. The human liver clears both asymmetric and symmetric dimethylarginine. *Hepatology* 2005;41(3):559-65.
16. Banjarnahor S, Rodionov RN, König J, Maas R. Transport of L-arginine related cardiovascular risk markers. *J Clin Med.* 2020;9(12):3975. doi: 10.3390/jcm9123975.
17. Kielstein JT, Bode-Böger SM, Frölich JC, Ritz E, Haller H, Fliser D. Asymmetric dimethylarginine, blood pressure, and renal perfusion in elderly subjects. *Circulation* 2003;107(14):1891-5.
18. Goonasekera CD, Rees DD, Woolard P, Frend A, Shah V, Dillon MJ. Nitric oxide synthase inhibitors and hypertension in children and adolescents. *J Hypertens.* 1997;15(8):901-9.
19. Wang D, Strandgaard S, Iversen J, Wilcox CS. Asymmetric dimethylarginine, oxidative stress, and vascular nitric oxide synthase in essential hypertension. *Am J Physiol Regul Integr Comp Physiol.* 2009;296(2):195-200.
20. Surdacki A, Nowicki M, Sandmann J, Tsikas D, Boeger RH, Bode-Boeger SM, et al. Reduced urinary excretion of nitric oxide metabolites and increased plasma levels of asymmetric dimethylarginine in men with essential hypertension. *J Cardiovasc Pharmacol.* 1999;33(4):652-8.
21. Usui M, Matsuoka H, Miyazaki H, Ueda S, Okuda S, Imaizumi T. Increased endogenous nitric oxide synthase inhibitor in patients with congestive heart failure. *Life Sci.* 1998;62(26):2425-30.
22. Feng Q, Lu X, Fortin AJ, Pettersson A, Hedner T, Kline RL, et al. Elevation of an endogenous inhibitor of nitric oxide synthesis in experimental congestive heart failure. *Cardiovasc Res.* 1998;37(3):667-75.
23. Saitoh M, Osanai T, Kamada T, Matsunaga T, Ishizaka H, Hanada H, et al. High plasma level of asymmetric dimethylarginine in patients with acutely exacerbated congestive heart failure: Role in reduction of plasma nitric oxide level. *Heart Vessels* 2003;18(4):177-82.
24. Achan V, Broadhead M, Malaki M, Whitley G, Leiper J, MacAllister R, et al. Asymmetric dimethylarginine causes hypertension and cardiac dysfunction in humans and is actively metabolized by dimethylarginine dimethylaminohydrolase. *Arterioscler Thromb Vasc Biol.* 2003;23(8):1455-9.
25. Kielstein JT, Impraïm B, Simmel S, Bode-Böger SM, Tsikas D, Frölich JC, et al. Cardiovascular effects of systemic nitric oxide synthase inhibition with asymmetrical dimethylarginine in humans. *Circulation* 2004;109(2):172-7.
26. MacAllister R, Vallance P. Nitric oxide in essential and renal hypertension. *J Am Soc Nephrol.* 1994;5(4):1057-65.
27. Kielstein JT, Simmel S, Bode-Böger SM, Roth HJ, Schmidt-Gayk H, Haller H, et al. Subpressor dose asymmetric dimethylarginine modulates renal function in humans through nitric oxide synthase inhibition. *Kidney Blood Press Res.* 2004;27(3):143-7.
28. Ruilope LM, Lahera V, Rodicio JL, Romero JC. Participation of nitric oxide in the regulation of renal function: possible role in the genesis of arterial hypertension. *J Hypertens.* 1994;12(6):625-31.
29. Bech JN, Nielsen CB, Pedersen EB. Effects of systemic NO synthesis inhibition on RPF, GFR, UNa, and vasoactive hormones in healthy humans. *Am J Physiol.* 1996;270(5 Pt 2):845-51.
30. Neiryck N, Vanholder R, Schepers E, Eloot S, Pletinck A, Glorieux G. An update on uremic toxins. *Int Urol Nephrol.* 2013;45(1):139-50.
31. Jacobi J, Tsao PS. Asymmetrical dimethylarginine in renal disease: Limits of variation or variation limits? A systematic review. *Am J Nephrol.* 2008;28(2):224-37.
32. Teerlink T. ADMA metabolism and clearance. *Vasc Med.* 2005;10(Suppl 1):73-81.
33. Palm F, Onozato ML, Luo Z, Wilcox CS. Dimethylarginine dimethylaminohydrolase (DDAH): Expression, regulation, and function in the cardiovascular and renal systems. *American Journal of Physiology-Heart and Circulatory Physiology* 2007;293(6):3227-45.
34. Chen Y, Li Y, Zhang P, Traverse JH, Hou M, Xu X, et al. Dimethylarginine dimethylaminohydrolase and endothelial dysfunction in failing hearts. *Am J Physiol Heart Circ Physiol.* 2005;289(5):2212-9.
35. Chen Y, Park S, Li Y, Missov E, Hou M, Han X, et al. Alterations of gene expression in failing myocardium following left ventricular assist device support. *Physiol Genomics* 2003;14(3):251-60.
36. Ito A, Tsao PS, Adimoolam S, Kimoto M, Ogawa T, Cooke JP. Novel mechanism for endothelial dysfunction: Dysregulation of dimethylarginine dimethylaminohydrolase. *Circulation* 1999;99(24):3092-5.
37. Lin KY, Ito A, Asagami T, Tsao PS, Adimoolam S, Kimoto M, et al. Impaired nitric oxide synthase pathway in diabetes mellitus: Role of asymmetric dimethylarginine and dimethylarginine dimethylaminohydrolase. *Circulation* 2002;106(8):987-92.
38. Valkonen VP, Tuomainen TP, Laaksonen R. DDAH gene and cardiovascular risk. *Vasc Med.* 2005;10(Suppl 1):45-8.
39. Tojo A, Welch WJ, Bremer V, Kimoto M, Kimura K, Omata M, et al. Colocalization of demethylating enzymes and NOS and functional effects of methylarginines in rat kidney. *Kidney Int.* 1997;52(6):1593-601.
40. Abhary S, Kasmeridis N, Burdon KP, Kuot A, Whiting MJ, Yew WP, et al. Diabetic retinopathy is associated with elevated serum asymmetric and symmetric dimethylarginines. *Diabetes Care* 2009;32(11):2084-6.
41. Stojanovic I, Djordjevic G, Pavlovic R, Djordjevic V, Pavlovic D, Cvetkovic T, et al. The importance of L-arginine metabolism modulation in diabetic patients with distal symmetric polyneuropathy. *J Neurol Sci.* 2013;324(1-2):40-4.
42. Hanai K, Babazono T, Nyumura I, Toya K, Tanaka N, Tanaka M, et al. Asymmetric dimethylarginine is closely associated with the development and progression of nephropathy in patients with type 2 diabetes. *Nephrol Dial Transplant.* 2009;24(6):1884-8.



43. Liu J, Li C, Chen W, He K, Ma H, Ma B, et al. Relationship between serum asymmetric dimethylarginine level and microvascular complications in diabetes mellitus: A meta-analysis. *Biomed Res Int.* 2019;2019:2941861.
44. Lee W, Lee HJ, Jang HB, Kim HJ, Ban HJ, Kim KY, et al. Asymmetric dimethylarginine (ADMA) is identified as a potential biomarker of insulin resistance in skeletal muscle. *Sci Rep.* 2018;8(1):2133.
45. Mookerjee RP, Dalton RN, Davies NA, Hodges SJ, Turner C, Williams R, et al. Inflammation is an important determinant of levels of the endogenous nitric oxide synthase inhibitor asymmetric dimethylarginine (ADMA) in acute liver failure. *Liver Transpl.* 2007;13(3):400-5.
46. Nijveldt RJ, Teerlink T, van Leeuwen PA. The asymmetrical dimethylarginine (ADMA)-multiple organ failure hypothesis. *Clin Nutr.* 2003;22(1):99-104.
47. Ferrigno A, Di Pasqua LG, Berardo C, Richelmi P, Vairetti M. Liver plays a central role in asymmetric dimethylarginine-mediated organ injury. *World journal of gastroenterology.* 2015;21(17):5131-7.
48. Mookerjee RP, Malaki M, Davies NA, Hodges SJ, Dalton RN, Turner C, et al. Increasing dimethylarginine levels are associated with adverse clinical outcome in severe alcoholic hepatitis. *Hepatology.* 2007;45(1):62-71.
49. Lluch P, Torondel B, Medina P, Segarra G, Del Olmo JA, Serra MA, et al. Plasma concentrations of nitric oxide and asymmetric dimethylarginine in human alcoholic cirrhosis. *J Hepatol.* 2004;41(1):55-9.
50. Davids M, Richir MC, Visser M, Ellger B, van den Berghe G, van Leeuwen PAM, et al. Role of dimethylarginine dimethylaminohydrolase activity in regulation of tissue and plasma concentrations of asymmetric dimethylarginine in an animal model of prolonged critical illness. *Metabolism.* 2012;61(4):482-90.
51. Khalil AA, Tsikas D, Akolekar R, Jordan J, Nicolaides KH. Asymmetric dimethylarginine, arginine and homoarginine at 11-13 weeks' gestation and preeclampsia: A case-control study. *J Hum Hypertens.* 2013;27(1):38-43.
52. Böger RH, Diemert A, Schwedhelm E, Lüneburg N, Maas R, Hecher K. The role of nitric oxide synthase inhibition by asymmetric dimethylarginine in the pathophysiology of preeclampsia. *Gynecol Obstet Invest.* 2010;69(1):1-13.
53. Savvidou MD, Hingorani AD, Tsikas D, Frölich JC, Vallance P, Nicolaides KH. Endothelial dysfunction and raised plasma concentrations of asymmetric dimethylarginine in pregnant women who subsequently develop pre-eclampsia. *Lancet.* 2003;361(9368):1511-7.
54. Speer PD, Powers RW, Frank MP, Harger G, Markovic N, Roberts JM. Elevated asymmetric dimethylarginine concentrations precede clinical preeclampsia, but not pregnancies with small-for-gestational-age infants. *Am J Obstet Gynecol.* 2008;198(1):112.e1-7.
55. Pettersson A, Hedner T, Milsom I. Increased circulating concentrations of asymmetric dimethyl arginine (ADMA), an endogenous inhibitor of nitric oxide synthesis, in preeclampsia. *Acta Obstet Gynecol Scand.* 1998;77(8):808-13.
56. Maas R, Böger RH, Schwedhelm E, Casas JP, López-Jaramillo P, Serrano N, et al. Plasma concentrations of asymmetric dimethylarginine (ADMA) in Colombian women with pre-eclampsia. *Jama.* 2004;291(7):823-4.
57. Maeda T, Yoshimura T, Okamura H. Asymmetric dimethylarginine, an endogenous inhibitor of nitric oxide synthase, in maternal and fetal circulation. *J Soc Gynecol Investig.* 2003;10(1):2-4.
58. Saarelainen H, Valtonen P, Punnonen K, Laitinen T, Raitakari OT, Juonala M, et al. Subtle changes in ADMA and L-arginine concentrations in normal pregnancies are unlikely to account for pregnancy-related increased flow-mediated dilatation. *Clin Physiol Funct Imaging.* 2008;28(2):120-4.
59. Braekke K, Ueland PM, Harsem NK, Staff AC. Asymmetric dimethylarginine in the maternal and fetal circulation in preeclampsia. *Pediatric Research* 2009;66(4):411-5.
60. Hannemann J, Balfanz P, Schwedhelm E, Hartmann B, Ule J, Müller-Wieland D, et al. Elevated serum SDMA and ADMA at hospital admission predict in-hospital mortality of COVID-19 patients. *Sci Rep.* 2021;11(1):9895.
61. Fang W, Jiang J, Su L, Shu T, Liu H, Lai S, et al. The role of NO in COVID-19 and potential therapeutic strategies. *Free Radic Biol Med.* 2021;163:153-62.
62. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al. Endothelial cell infection and endotheliitis in COVID-19. *The Lancet* 2020;395(10234):1417-8.
63. Ueda S, Kato S, Matsuoka H, Kimoto M, Okuda S, Morimatsu M, et al. Regulation of cytokine-induced nitric oxide synthase by asymmetric dimethylarginine: Role of dimethylarginine dimethylaminohydrolase. *Circ Res.* 2003;92(2):226-33.
64. Hang do TT, Song JY, Kim MY, Park JW, Shin YK. Involvement of NF- κ B in changes of IFN- γ -induced CIITA/MHC-II and iNOS expression by influenza virus in macrophages. *Mol Immunol.* 2011;48(9-10):1253-62.
65. Liu X, Jana M, Dasgupta S, Koka S, He J, Wood C, et al. Human immunodeficiency virus type 1 (HIV-1) tat induces nitric-oxide synthase in human astroglia. *J Biol Chem.* 2002;277(42):39312-9.
66. Chen L, Hsieh MS, Ho HC, Liu YH, Chou DT, Tsai SH. Stimulation of inducible nitric oxide synthase by monosodium urate crystals in macrophages and expression of iNOS in gouty arthritis. *Nitric Oxide* 2004;11(3):228-36.
67. Chang PC, Chen TH, Chang CJ, Hou CC, Chan P, Lee HM. Advanced glycosylation end products induce inducible nitric oxide synthase (iNOS) expression via a p38 MAPK-dependent pathway. *Kidney Int.* 2004;65(5):1664-75.
68. Cho HJ, Xie QW, Calaycay J, Mumford RA, Swiderek KM, Lee TD, et al. Calmodulin is a subunit of nitric oxide synthase from macrophages. *J Exp Med.* 1992;176(2):599-604.
69. Zhao K, Huang Z, Lu H, Zhou J, Wei T. Induction of inducible nitric oxide synthase increases the production of reactive oxygen species in RAW264.7 macrophages. *Biosci Rep.* 2010;30(4):233-41.
70. Sharma JN, Al-Omran A, Parvathy SS. Role of nitric oxide in inflammatory diseases. *Inflammopharmacology* 2007;15(6):252-9.
71. Dominic P, Ahmad J, Bhandari R, Pardue S, Solorzano J, Jaisingh K, et al. Decreased availability of nitric oxide and hydrogen sulfide is a hallmark of COVID-19. *Redox Biol.* 2021;43:101982.
72. Green SJ. Covid-19 accelerates endothelial dysfunction and nitric oxide deficiency. *Microbes Infect.* 2020;22(4-5):149-50.
73. Sydow K, Münzel T. ADMA and oxidative stress. *Atheroscler Suppl.* 2003;4(4):41-51.
74. Hadi HA, Carr CS, Al Suwaidi J. Endothelial dysfunction: Cardiovascular risk factors, therapy, and outcome. *Vasc Health Risk Manag.* 2005;1(3):183-98.
75. Cooke JP. Does ADMA cause endothelial dysfunction? *Arterioscler Thromb Vasc Biol.* 2000;20(9):2032-7.
76. Förstermann U, Sessa WC. Nitric oxide synthases: regulation and function. *Eur Heart J.* 2012;33(7):829-37, 37a-37d.
77. Gardiner SM, Kemp PA, Bennett T, Palmer RM, Moncada S. Regional and cardiac haemodynamic effects of NG, NG-dimethyl-L-arginine and their reversibility by vasodilators in conscious rats. *Br J Pharmacol.* 1993;110(4):1457-64.



78. Calver A, Collier J, Leone A, Moncada S, Vallance P. Effect of local intra-arterial asymmetric dimethylarginine (ADMA) on the forearm arteriolar bed of healthy volunteers. *J Hum Hypertens*. 1993;7(2):193-4.
79. Tsikas D, Böger RH, Sandmann J, Bode-Böger SM, Frölich JC. Endogenous nitric oxide synthase inhibitors are responsible for the L-arginine paradox. *FEBS Letters*. 2000;478(1):1-3.
80. Arancibia-Garavilla Y, Toledo F, Casanello P, Sobrevia L. Nitric oxide synthesis requires activity of the cationic and neutral amino acid transport system y+L in human umbilical vein endothelium. *Exp Physiol*. 2003;88(6):699-710.
81. Németh B, Ajtay Z, Hejzel L, Ferenci T, Ábrám Z, Murányi E, et al. The issue of plasma asymmetric dimethylarginine reference range – A systematic review and meta-analysis. *PLOS ONE* 2017;12(5):e0177493.
82. Schulze F, Lenzen H, Hanefeld C, Bartling A, Osterziel KJ, Goudeva L, et al. Asymmetric dimethylarginine is an independent risk factor for coronary heart disease: results from the multicenter Coronary Artery Risk Determination investigating the Influence of ADMA Concentration (CARDIAC) study. *Am Heart J*. 2006;152(3):493.e1-8.
83. Horowitz JD, Heresztyn T. An overview of plasma concentrations of asymmetric dimethylarginine (ADMA) in health and disease and in clinical studies: methodological considerations. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2007;851(1-2):42-50.
84. Sydow K, Fortmann SP, Fair JM, Varady A, Hlatky MA, Go AS, et al. Distribution of asymmetric dimethylarginine among 980 healthy, older adults of different ethnicities. *Clin Chem*. 2010;56(1):111-20.
85. Schwedhelm E. Quantification of ADMA: Analytical approaches. *Vasc Med*. 2005;10(Suppl 1):89-95.
86. Tsikas D. A critical review and discussion of analytical methods in the L-arginine/nitric oxide area of basic and clinical research. *Anal Biochem*. 2008;379(2):139-63.
87. Weaving G, Rocks BF, Bailey MP, Titheradge MA. Arginine and methylated arginines in human plasma and urine measured by tandem mass spectrometry without the need for chromatography or sample derivatisation. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2008;874(1-2):27-32.
88. Martens-Lobenhoffer J, Bode-Böger SM. Chromatographic-mass spectrometric methods for the quantification of L-arginine and its methylated metabolites in biological fluids. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2007;851(1-2):30-41.
89. Teerlink T, Luo Z, Palm F, Wilcox CS. Cellular ADMA: Regulation and action. *Pharmacological Research* 2009;60(6):448-60.
90. Bedford MT, Clarke SG. Protein arginine methylation in mammals: Who, what, and why. *Mol Cell*. 2009;33(1):1-13.
91. Thiebaut C, Eve L, Poulard C, Le Romancer M. Structure, activity, and function of PRMT1. *Life* 2021;11(11):1147.
92. Dillon MB, Bachovchin DA, Brown SJ, Finn MG, Rosen H, Cravatt BF, et al. Novel inhibitors for PRMT1 discovered by high-throughput screening using activity-based fluorescence polarization. *ACS Chem Biol*. 2012;7(7):1198-204.
93. Yin QF, Fu SH, He P, Xiong Y. Dimethylarginine dimethylaminohydrolase inhibition and asymmetric dimethylarginine accumulation contribute to endothelial dysfunction in rats exposed to glycosylated protein: Effects of aminoguanidine. *Atherosclerosis* 2007;190(1):53-61.
94. Wakino S, Hayashi K, Tatematsu S, Hasegawa K, Takamatsu I, Kanda T, et al. Pioglitazone lowers systemic asymmetric dimethylarginine by inducing dimethylarginine dimethylaminohydrolase in rats. *Hypertension Research* 2005;28(3):255-62.
95. Yin QF, Xiong Y. Pravastatin restores DDAH activity and endothelium-dependent relaxation of rat aorta after exposure to glycated protein. *Journal of Cardiovascular Pharmacology* 2005;45(6):525-32.
96. Jiang JL, Zhang XH, Li NS, Rang WQ, Feng Y, Hu CP, et al. Probuco decreases asymmetrical dimethylarginine level by alternation of protein arginine methyltransferase I and dimethylarginine dimethylaminohydrolase activity. *Cardiovasc Drugs Ther*. 2006;20(4):281-94.
97. Hu T, Chouinard M, Cox AL, Sipes P, Marcelo M, Ficorilli J, et al. Farnesoid X receptor agonist reduces serum asymmetric dimethylarginine levels through hepatic dimethylarginine dimethylaminohydrolase-1 gene regulation*. *Journal of Biological Chemistry* 2006;281(52):39831-8.
98. Tain YL, Huang LT, Lin IC, Lau YT, Lin CY. Melatonin prevents hypertension and increased asymmetric dimethylarginine in young spontaneous hypertensive rats. *J Pineal Res*. 2010;49(4):390-8.
99. Yang ZC, Wang KS, Wu Y, Zou XQ, Xiang YY, Chen XP, et al. Asymmetric dimethylarginine impairs glucose utilization via ROS/TLR4 pathway in adipocytes: An effect prevented by vitamin E. *Cell Physiol Biochem*. 2009;24(1-2):115-24.
100. Lee Y, Mehrotra P, Basile D, Ullah M, Singh A, Skill N, et al. Specific lowering of asymmetric dimethylarginine by pharmacological dimethylarginine dimethylaminohydrolase improves endothelial function, reduces blood pressure and ischemia-reperfusion injury. *J Pharmacol Exp Ther*. 2021;376(2):181-9.
101. Strobel J, Mieth M, Endress B, Auge D, König J, Fromm MF, et al. Interaction of the cardiovascular risk marker asymmetric dimethylarginine (ADMA) with the human cationic amino acid transporter 1 (CAT1). *J Mol Cell Cardiol*. 2012;53(3):392-400.
102. Banjarnahor S, König J, Maas R. Screening of commonly prescribed drugs for effects on the CAT1-mediated transport of L-arginine and arginine derivatives. *Amino Acids* 2022;54(7):1101-8.
103. Maas R. Pharmacotherapies and their influence on asymmetric dimethylarginine (ADMA). *Vasc Med*. 2005;10(Suppl 1):49-57.
104. Yang TL, Chen MF, Xia X, Luo BL, Li YJ. Effect of fenofibrate on the level of asymmetric dimethylarginine in individuals with hypertriglyceridemia. *Eur J Clin Pharmacol*. 2006;62(3):179-84.
105. Holven KB, Haugstad TS, Holm T, Aukrust P, Ose L, Nenseter MS. Folic acid treatment reduces elevated plasma levels of asymmetric dimethylarginine in hyperhomocysteinaemic subjects. *Br J Nutr*. 2003;89(3):359-63.
106. Chang JW, Lee EK, Kim TH, Min WK, Chun S, Lee KU, et al. Effects of alpha-lipoic acid on the plasma levels of asymmetric dimethylarginine in diabetic end-stage renal disease patients on hemodialysis: A pilot study. *Am J Nephrol*. 2007;27(1):70-4.